The effects of predation risk, shelter, and food availability on the reproduction of Aegean Wall lizards (*Podarcis erhardii*) in the Greek islands

Yilun Zhao

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Advisor: Associate Professor Johannes Foufopoulos Lecturer Shannon Brines

ABSTRACT

Reproductive investment, including the average number of offspring produced by an organism, is one of the fundamental characteristics of a species. Among other things, it predicts a species' resilience to environmental disruption: taxa that produce more offspring are able to recover more quickly from environmental perturbations and survive long-term environmental change. Despite the clear importance of this trait, ecologists do not have a good understanding of the primary drivers shaping the reproductive investment of each species. To answer this question, I compare the reproductive efforts of numerous island populations of the Aegean Wall Lizard (Podarcis erhardii), which differ in multiple key environmental characteristics. I test three hypotheses, namely that reproductive investment (measured as clutch size, clutch volume and egg volume) is: 1) positively associated with predation risk ['Predation Risk Hypothesis']; 2) positively associated with the presence of reliable vegetation cover that provides shelter ['Gravid Female Protection Hypothesis'], and 3) limited by (and hence positively correlated with) food availability ['Food Limitation Hypothesis']. Although field data are consistent with all three hypotheses, statistical analysis shows strong support for the Predation Risk Hypothesis. The result not only shed light on which fundamental forces shape reproductive investment in island vertebrates, but can also help set conservation priorities by identifying the most sensitive populations because reduced reproductive ability can be predicted based on easily quantifiable island characteristics (number of sympatric predator species).

Keywords: reproduction, reptile, conservation, island, predators, food.

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ABSTRACTI
ACKNOWLEDGEMENTS II
INTRODUCTION
METHODS
STUDY SITES
STUDY ORGANISM
Reproductive Traits
PREDATION PRESSURE
MEASUREMENT OF FOOD AVAILABILITY
MEASUREMENT OF VEGETATION
STATISTICAL ANALYSES
RESULTS
LINEAR MODELS OF ISLAND SIZE AND INDEPENDENT VARIABLES
CORRELATIONS
EFFECT OF PREDATOR RICHNESS, BIOMASS OF ARTHROPODS, AND NDVI ON REPRODUCTIVE
TRAITS
Hypothesis Testing
TABLES7
FIGURES
DISCUSSION
BIBLIOGRAPHY
APPENDICES

Table of Contents

INTRODUCTION

Island endemics constitute a significant component of earth's biodiversity. Despite the relatively small area they inhabit, these taxa also represent a disproportionate fraction of the planets endangered biodiversity, with fully 41% of the world's endangered terrestrial species found predominately in island systems (Spatz et al., 2017). Island species face particularly high extinction rates (Case, Bolger, & Richman, 1992) as they are impacted by numerous threats including invasive species (Bellard, Cassey, & Blackburn, 2016; Wikelski et al., 2004) and global climate change (Foufopoulos, Kilpatrick, & Ives, 2011). The rapidly deteriorating conservation status of island vertebrates adds urgency to understanding the endogenous and exogenous factors that drive their decline.

Island vertebrates are often characterized by a distinct suite of life history changes referred to under the umbrella term 'island syndrome' (Adler & Levins, 1994). These changes have been observed across a broad range of disparate island organisms including mammals (Goltsman, et al., 2005), birds (Clegg & Owens, 2002), and reptiles (Novosolov & Meiri, 2013). Among reptiles, typical changes associated with the island syndrome include shifts in body size (Case, 1978; Meiri, 2007; Pafilis et al., 2011), modified limb length and head shapes (Herrel et al., 2008; Runemark, Hansson, Pafilis, Valakos, & Svensson, 2010), as well as longer life spans and lower growth rates (Andrews, 1976). Island lizards tend to achieve higher densities (Case, 1975; Novosolov et al., 2016) and may also display altered levels of intraspecific aggressiveness (Pafilis, Meiri, Foufopoulos, & Valakos, 2009), as well as attenuated morphological anti-predator behaviors (Brock, Bednekoff, Pafilis, & Four four poulos, 2015a; Li et al., 2011; Stamps & Buechner, 1985) and adaptations (Pafilis et al., 2009). Probably the most widely recognized aspect of island syndrome is the altered pattern of reproductive investment, typically in the form of smaller clutches of larger eggs (Huang, 2007;-Smith & Fretwell, 1974; Blondel, 2000). However, this pattern is neither universal (Galán, 2003) nor consistent (Novosolov et al. 2012), and it is not clear which factors drive reproductive investment.

Reproductive investment is a central aspect of a species' life history and can vary tremendously even across closely related taxa (Du et al., 2005; Mesquita et al., 2016). Studies over the last half century have revealed the multitude of factors that shape reproduction, including climate (Angilletta et al., 2004; Roitberg et al., 2013), latitude (Sinervo, 1990; Mesquitea et al., 2016), elevation (Ballinger, 1979; Cruz-Elizalde & Ramírez-Bautista, 2016), foraging mode (Vitt & Congdon, 1978), phylogenetic history (e.g. Mesquita & Colli, 2010; Werneck et al., 2009) and general body bauplan (Roitberg et al., 2013), but also proximate environmental conditions such as refugium shape (Vitt, 1981; Goodman et al., 2009), resource availability (Ballinger, 1977), and infection with parasites (Foufopoulos, 1999).

Chief among others, reproductive effort is thought to be subject to the constraints imposed by resource availability (Ballinger, 1977). Numerous studies have demonstrated the importance of both condition and seasonal nutrient intake in shaping both clutch size and clutch mass (Jordan & Snell, 2002; Hoy et al., 2016; Warne et al., 2012; Pafilis et al., 2011). In practice, comparisons made between high and lower quality habitats, or between high and low rainfall years have revealed that food availability can, but does not have to be, a driver of reproductive investment (Ballinger, 1977; Du, 2006). Moreover, it is not clear to which extend food availability modulates individual clutch size across years, and between

individuals of a population, or whether it acts as a long-term driver of macroevolutionary differences in reproductive output across different populations of a species.

Beyond resource availability, perhaps the most attention has been paid to the role of predation on reproductive investment. Both theoretical and empirical studies have revealed that predator-caused mortality can be a particularly important driver of an organism's patterns of reproduction (Schaffer, 1974; Vitt & Congdon, 1978; Stearns, 1977; 1992; Magnhagen, 1991). Indeed, according to classic life history theory, in areas of high mortality, selection should favor early onset of reproduction and high reproductive investment, even if that comes at the cost of future reproduction (Schwarzkopf, 1994). Nonetheless, whether predator-induced mortality affects reproductive investment in relatively isolated island populations have never been tested in a rigorous manner.

Numerous studies have shown that increases in reproductive investment in females come at the cost of declining running ability (VanDamme et al., 1989) and that this decline can also undermine longer-term survival (Mils et al., 2000) revealing a trade-off between current and future reproduction (Reznick, 1985; 1992). Gravid females can partially compensate for some of this loss in escape ability by modifying their foraging behavior (Cooper et al., 1990; Braña, 1993; Bauwens & Thoen, 1981) and initiating escape activities earlier (Brown & Shine, 2004; Lima & Dill, 1990; Magnhagen, 1991). Furthermore, gravid females stay closer to refugia (Braña, 1993; Bauwens & Thoen, 1981) and reduce their visibility through appropriate microhabitat selection (Cooper et al., 1990). Consequently, habitats that lack, or have only insufficient cover and refugia, may exert selective pressure on reduced reproductive investment (Cruz-Elizalde et al., 2017; Haenel, 2011). For example, in Platynotus semitaeniatus, an iguanid lizard species that seeks refuge in narrow rock crevices, females have evolved smaller clutches and reduced clutch mass which in turn allow them to squeeze into smaller crevices to avoid predation (Vitt, 1981; Goodman et al., 2009). Nonetheless, beyond this special case, the hypothesis that availability of cover can affect reproductive investment has, to our knowledge, never been explicitly tested.

While a steadily growing body of literature has described patterns of vertebrate reproduction on islands, very few studies have investigated which are the proximate drivers that have shaped the evolution of these patterns. Because so many co-varying factors have the potential to affect reproductive output in wildlife (Dunham & Miles, 1985) an increasing number of studies have taken an intraspecific approach and compared populations across a single widespread species (Niewiarowski et al., 2004; Díaz et al., 2012; Iverson et al., 1997; Roitberg et al., 2013). By comparing multiple locations within the circumscribed geographic range of a single species, investigators can control for many confounding factors such as climate, elevation, body architecture and phylogenetic effects and identify which microevolutionary processes are ultimately responsible for the observed variation in reproductive output. Focusing on island populations has the added benefit that they are evolutionarily discreet and, especially if small enough, can be considered homogenous in terms of prevailing environmental conditions.

In this study we describe and analyze the evolutionary patterns of reproductive investment in the Aegean Wall Lizard (*Podarcis erhardii*) - a lacertid species widely distributed across the Greek archipelago. By taking advantage of an unusual ecological setting where pronounced life-history differences exist between numerous well-characterized island populations both in reproductive investment and in the prevailing ecological

conditions, we test three fundamental hypotheses regarding the forces driving the evolution of reproductive effort in island taxa. Specifically, we test whether clutch size and clutch volume are determined by:

- (1) amount of food available to the lizards (Food Limitation Hypothesis),
- (2) amount of shelters available to the lizards, and (Gravid Female Protection Hypothesis)
- (3) species richness of the local predator community (Predation Risk Hypothesis).

By elucidating these relationships, we not only shed light on the drivers of the island syndrome but also help understand the fundamental causes of reproductive investment in ectothermic organisms.

METHODS

Study Sites

The study region lies predominantly in the Aegean Sea archipelago located between the southern Balkan peninsula in the west and the Anatolian mainland in the east. The study was conducted on 12 Aegean islands - 8 islands in the Cyclades Cluster and 4 islands in the Sporades Cluster, as well as 2 sites located in nearby mainland Greece (Figure 1). Island size ranges from 0.005 km² to 429 km². The climate of the region is typical Mediterranean, with long, dry and warm summers and mild, rainy winters (Giorgi & Lionello, 2008). The vegetation cover on the study sites consists mostly of xerophytic, summer-deciduous, coastal heaths termed 'phrygana' (comprised of diverse, spinose plant communities rich in aromatic taxa) as well as of agricultural fields, and sclerophyllous evergreen maquis (Fielding & Turland, 2008). The vegetation has been shaped by millennia of anthropogenic human activities including terraced agriculture and small ruminant grazing (Grove & Rackham, 2003).

Study Organism

The Aegean Wall Lizard (*Podarcis erhardii*) is a medium-sized lacertid lizard species with an adult SVL of 49-78 mm (Valakos et al., 2008). This species is widely distributed across the Greek mainland and the Aegean Sea islands (Brock et al., 2015). The species usually mates in spring and females lay their eggs in the period from April to July. Depending on the local conditions, eggs are hatched in middle to late summer (Valakos et al, 2008). Aegean wall lizards occur in a wide range of open habitats, with a preference for open, stony regions and tend to be absent from areas with dense vegetation and closed forest cover. They are particularly common in areas that provide refugia in the form of broken-up terrain and anthropogenic structures such as dry-stone walls and terraces (Brock et al., 2015b). P. erhardii consume a broad range of arthropods (Arnold, 1987), with a particular emphasis on Coleoptera, Orthoptera, and soft-bodied larvae (Adamopoulou, Valakos & Pafilis 1999). Additional secondary food items include snails (Adamopoulou, Valakos & Pafilis 1999) and even fruit (Brock, Donihue, & Pafilis, 2014), although the species is not known to consume nectar or vegetable matter like other island taxa. Recent observations suggest that the species displays at least occasionally cannibalistic tendencies (Adamopoulou, Valakos & Pafilis, 1999).

Reproductive Traits

Morphological traits were obtained from museum specimens. Snout-vent length (SVL) and reproductive traits, including clutch size, egg volume, and clutch volume were collected from specimens deposited at the Zoological Museum of Bonn (Germany) as well as from the Zoologische Staatssammlung in Munich (Germany). Following dissection, the number of oviductal eggs were recorded as clutch size. Size of oviductal eggs was recorded using digital calipers including the longest and shortest axes. Egg volume was determined by using the equation for volume (V) of an ellipsoid:

$$V = \frac{4}{3}\pi\alpha b^2$$

where α is half of the longest axis, and b is half of the shortest axis. Clutch volume for each female was calculated as the sum of the individual egg volumes (Mayhew, 1963).

Predation Pressure

Numerous types of predators feed on Aegean wall lizards, but their individual presence varies greatly across the range of *P. erhardii*, depending on ecological, vicariance and biogeographic factors. While mainland sites tend to harbor diverse communities of avian, mammalian, and reptile predators, some of the smallest islands fail to support even a single type of predator, hence creating a wide range of predation regimes. To obtain a more formal estimation of the predation environment that each population experiences, we follow the methodology of previous authors (Pafilis et al., 2009; Brock et al., 2015) and use predator species richness as a proxy of predation risk in each area. Predator presence data were obtained from the published literature and was often confirmed from our own field observations.

Measurement of Food Availability

We investigated the diet of *P. erhardii* across the study sites spanning a range of ecological conditions by studying biomass and abundance of arthropods in these sites during the key period during which females forage to form their clutches. From May to July 2017, 5 pitfall and sticky trap pairs were set on each island to collect crawling and flying arthropods. All pitfalls and sticky traps were set in randomly-selected areas with natural vegetation in the immediate vicinity of the sites where reproductive data specimens were collected. Crawling arthropods populations were analyzed by deploying five 400ml pitfalls containing antifreeze. To determine flying insect populations, we set up five 15.24cm x 30.48cm sticky traps placed on 30cm stakes over, or near, the pitfalls on each island. Following collection, all arthropods from pitfalls were washed with ethanol to remove dirt and antifreeze and stored in a 2oz. plastic wide-mouth jars for subsequent identification and measurement. All arthropods were identified to order and length was measured to the nearest *mm* using a ruler. Approximate biomass of each individual was then calculated using the length-to-biomass equation (Rogers, Hinds, & Buschbom, 1976):

$W = 0.0305 \cdot L^{2.62}$

The abundance and biomass of each sticky trap or pitfall trap was calculated and each island's abundance and biomass were calculated by averaging each sticky strip or pitfall. Because the deployment time of sticky strips and pitfalls from each island was different, the average abundance and biomass was standardized on a 48-hour basis.

Measurement of Vegetation

We measured vegetation cover by utilizing a Normalized Difference Vegetation Index (NDVI) ("Measuring Vegetation". NASA Earth Observatory.). For each island, we downloaded Landsat 8 OLI/ TIRS level 2 (surface reflectance) images from Earth Explorer, U.S. Geological Survey. To ensure accuracy of NDVI, we used only images with less than 10% cloud cover. NDVI of each surface reflectance image was calculated in Erdas Imagine 2016. Each image was then input in ArcGIS 10, and clipped to the sample area with 62.5m radius around sample site based on the size of the smallest of our field sites (Kokkinonisi). The NDVI of each pixel in sample area was exported and for each island, the NDVI of sample area was calculated by averaging the values of the corresponding pixels.

Statistical Analyses

Former studies show that island size influences vegetation, arthropods and predation risk (Kohn and Walsh, 1994; Shure and Phillips, 1991; Polis and Hurd, 1995). To test this hypothesis, we used linear models to find the relationship between island size (island area) and the amount of shelters (vegetation), food availability (biomass of arthropods) and predation risk (predator richness). Since island area has a highly skewed distribution, we used both island area and log-transformed island area as the independent variable when building linear regression models.

Because previous studies have shown that maternal SVL may affect reproductive traits, we included maternal SVL in some of the models as a covariate (Abell, Reyes, & Wiernasz, 1999; Pafilis et al., 2011). To avoid issues of collinearity we tested the correlations between all of the driving variables considered in this paper (maternal SVL, NDVI, predator richness, and biomass of arthropods) and excluded any complex models that had variables that were highly correlated with each other (r>0.5). Since multiple lizards from each island were measured and individuals from the same island are assumed to be more similar with each other, we built linear models and linear mixed effect models to check the effect of shelter amount, predation risk, and food availability on reproductive traits.

Afterwards, Akaike information criterion (AIC) was used for comparison of the models. For each reproductive trait, same type of models (linear model or linear mixed effect model) were compared by AIC. Independent variables in the model with the lowest AIC value were considered to be the most important factor affecting reproductive traits.

RESULTS

Linear Models of Island Size and Independent Variables

Results of linear models show that log-transformed island area has a positive relationship with log-transformed predator richness (r = 0.190, p<0.001, $R_{adj}^2=0.942$) (Figure 2). The log-transformed biomass of arthropods has a positive correlation with log-transformed island area (r = 0.152, p=0.0018, $R_{adj}^2=0.531$) (Figure 3). NDVI shows a positive relationship with island area (r = 0.0002752, p=0.023, $R_{adj}^2=0.308$) (Figure 4). Hence larger islands tend to have higher predation risk, food availability, and extend of shelter-providing vegetation cover (Appendix 1).

Correlations

The correlation between predator richness and NDVI is 0.650 (p<0.001), the correlation between predator richness and biomass of arthropods is 0.701 (p<0.001), and the correlation between arthropods biomass and NDVI is 0.600 (p<0.001). Because the correlations among predator richness, biomass of arthropods, and NDVI are all higher than 0.5 resulting in potential collinearity issues, we did not include them as independent variables in the same models during the model building process. The correlation between maternal snout-vent length and predator richness is 0.195 (p=0.002); the correlation between maternal snout-vent length and NDVI is 0.102 (p=0.111); the correlation between maternal snout-vent length and one of other three independent variables were included in same models.

Effect of Predator Richness, Biomass of Arthropods, and NDVI on Reproductive Traits

Among three reproductive traits, clutch size was found significantly related to predator richness (r = 0.087, P<0.001, R^2_{adj} = 0.173), biomass of arthropods (r = 0.0034153, P<0.001, R^2_{adj} = 0.07464) and NDVI (r = 1.7898, P<0.001, R^2_{adj} = 0.06778). Additionally, clutch volume has significant positive relationships with predator richness (r = 44.52, P<0.001, R^2_{adj} = 0.0673) and biomass of arthropods (r = 1.659, P = 0.00828, R^2_{adj} = 0.02424). We also found significant positive effects from maternal snout-vent length on clutch size (r = 0.06146, P<0.001, R^2_{adj} = 0.08326) and clutch volume (r = 30.07, P = 0.00505, R^2_{adj} = 0.0278). (Appendices 2, 3, 4)

Hypothesis Testing

Results of AIC comparisons are listed in Table 2, 3, 4, and 5 (Table 4 and Table 5 are in Appendix 6). Among all linear models and linear mixed effect models for explaining clutch size, the linear model contains predator richness and maternal SVL has the lowest AIC value (Table 2). For clutch volume, the linear mixed effect model including predator richness and maternal SVL as fixed terms, and location as the random term has the lowest AIC value, in terms of the highest explanation power (Table 3) although an alternative model that included NDVI, maternal SVL and location was only marginally worse (Appendix 5). Figure 5 and Figure 6 are the visualization of these two best models.

TABLES

 Table 1. Summary Table including island name, coordinates, island size, predator richness, biomass of arthropods, NDVI, clutch size and clutch volume.

Island Name	Coordinates	Island Size (km ²)	Predator Richness	Biomass of Arthropods (mg)	NDVI	Clutch Size	Clutch Volume (mm ³)
Kokkinonisi	39°9'38.757"N, 23°54'7.129"E	0.005	2	16.734	0.387	1.7±0.3(10)	598±234(10)
Mikropsathoura (Mougia)	39°28'56.262"N, 24°10'51.691"E	0.014	2	20.724	0.511	1.6±0.2(11)	219±60(11)
Agios Ioannis	36°36'36.327"N, 24°57'23.118"E	0.033	2	125.727	0.108	1.4±0.2(10)	433±143(10)
Kopria	36°59'27.899"N, 25°38'14.122"E	0.138	2	100.870	0.164	1.6±0.2(10)	805±254(10)
Glaronisi	36°55'15.371"N, 25°36'15.286"E	0.188	2	57.548	0.200	1.9±0.1(8)	362±100(8)
Ano Koufonisi	36°56'49.45"N, 25°36'20.237"E	5.770	5	96.874	0.237	2.2±0.2(9)	386±120(9)
Gioura	39°23'46.899"N, 24°10'20.407"E	11.052	6	191.591	0.311	1.8±0.3(9)	549±149(9)
Santorini	36°22'59.326"N, 25°28'29.843"E	76.197	9	106.454	0.312	1.7±0.1(24)	612±121(24)
Skopelos	39°7'30.145"N, 23°39'10.323"E	96.229	11	333.154	0.542	2.5±0.4(12)	947±235(12)
Tinos	37°33'31.293"N, 25°7'40.568"E	194.500	12	77.232	0.213	2.7±0.2(23)	1268±269(23)
Andros	37°53'23.97"N, 24°43'25.309"E	380.000	13	286.327	0.363	2.2±0.4(9)	777±289(9)
Naxos	37°4'54.364"N, 25°29'16.147"E	429.785	11	121.837	0.289	2.4±0.1(42)	1008±139(42)
Olympiada	39°59'45.907"N, 22°14'0.477"E	1000.000	16	278.560	0.485	2.9±0.2(35)	1258±175(35)
Vevi	40°46'27.065"N, 21°36'53.896"E	1000.000	18	229.523	0.682	2.9±0.2(34)	951±165(34)

Model	AICc	Δ AICc	Akaike Weight
$CS \sim P + SVL$	695.691	0.000	0.998
$CS \sim P$	708.225	12.533	1.895x10 ⁻³
$CS \sim B + SVL$	712.715	17.024	2.007x10 ⁻⁴
$CS \sim NDVI + SVL$	719.651	23.959	6.257x10 ⁻⁶
$CS \sim SVL$	733.504	37.812	6.141x10 ⁻⁹
$CS \sim B$	735.805	40.114	1.943x10 ⁻⁹
$CS \sim NDVI$	737.624	41.933	7.826x10 ⁻¹⁰

Table 2. AIC for seven linear models constructed to explain clutch size

 $\overline{\text{CS}}$ = Clutch Size, P = Predator Richness, B = Biomass of Arthropods, SVL = maternal snoutvent length. Models were compared with their AICc values and their associated Akaike weight which is calculated by the equation:

 $W_i(AIC) = exp(-0.5\Delta_i(AIC)) / \sum_{k=1}^k exp(-0.5\Delta_k(AIC)) \text{ (Turkheimer et al. 2003).}$

Model	AIC	ΔΑΙΟ	Akaike Weight
			8
CV ~ P + SVL + (1 Location)	4008.389	0	0.587
$CV \sim NDVI + SVL + (1 Location)$	4009.153	0.764	0.401
$CV \sim P + (1 Location)$	4017.407	9.018	6.461x10 ⁻³
$CV \sim B+SVL + (1 Location)$	4018.701	10.312	3.383x10 ⁻³
$CV \sim NDVI + (1 Location)$	4019.651	11.262	2.104x10 ⁻³
$CV \sim SVL + (1 Location)$	4022.031	13.642	6.401x10 ⁻⁴
$CV \sim B + (1 Location)$	4029.952	21.563	1.220x10 ⁻⁵

 $\overline{\text{CV} = \text{Clutch Volume}}$, $\overline{\text{P} = \text{Predator Richness}}$, $\overline{\text{B} = \text{Biomass of Arthropods}}$, $\overline{\text{SVL} = \text{maternal}}$ snout-vent length. Models were compared with their AICc values and their associated Akaike weight which is calculated by the equation:

 $W_i(AIC) = exp(-0.5\Delta_i(AIC)) / \sum_{k=1}^{k} exp(-0.5\Delta_k(AIC))$ (Turkheimer et al. 2003).

FIGURES



Figure 1. Map of the general area with the study sites marked in green circles.



Figure 2. Linear regression between log-transformed Island Area and log-transformed Predator Richness. Each circle represents an island. Log-transformed predator richness has a positive relationship with log-transformed island area.



Figure 3. Linear regression between log-transformed Island Area and log-transformed Biomass of arthropods. Each circle represents an island. Log-transformed biomass of arthropods has a positive relationship with log-transformed island area.



Figure 4. Linear regression between island area and NDVI. Each circle represents an island. NDVI has a positive relationship with island area.



Figure 5. Lizard clutch size against predator species richness and maternal body size. Each dot represents one lizard clutch, and wire mesh represents the predictions of the best model as provided from the AIC model comparison. x: Predator Species Richness, y: Maternal snout-vent length (in *mm*), z: clutch size (number of eggs).



Figure 6. Lizard clutch volume against predator species richness and maternal body size (in *mm*). Each dot represents one lizard clutch, and wire mesh represents the predictions of linear model. x: Predator species richness, y: Maternal snout-vent length (in *mm*), z: Clutch volume (in mm^3).

DISCUSSION

Reproductive investment is a fundamental component of a species' biology and has been the focus of many fruitful life-history studies (Tinkle, 1969; Tinkle et al., 1970; Stearns, 1992). It is also of practical importance: for example, small clutch size has been shown to be a critical predictor of vulnerability to extinction in lizards (Siliceo & Díaz, 2010). Reptiles have emerged as particularly useful study systems to investigate the evolution of different reproductive investments because of the tremendous variety in reproductive modes that are made possible by ectothermy (Shine, 2005; Mesquita et al., 2016). Because reproduction entails multiple conflicting demands and organisms operate under limited resources, (e.g. nutrients -Ballinger, 1977; maternal body cavity volume - Qualls and Shine, 1995) it is impossible for an individual to optimize all aspects of its life history (Mesquita et al., 2016). This in turn creates important trade-offs such as present-season versus future-season reproduction (Shine, 2003; Schaffer, 1974; Schwarzkopf, 1994; Miles et al., 2000; Shine & Schwarzkopf, 1992; Olsson et al., 2001). Other well-recognized trade-offs exist within a single clutch e.g. the fundamental choice between number of offspring and size of individual offspring (Sinervo, 1990; Sinervo and Licht, 1991). Ultimately, a lot of variation exists between, as well as within species (e.g. Angilletta et al., 2004) and much of it remains unexplained.

In this study we tested the effect of different factors on three different reproductive traits, clutch size, clutch volume and egg volume. We found that while clutch size was significantly and positively correlated with predation risk, food availability, and vegetation amount, the latter two of these relationships were weaker than the first one and had little explanatory power. Similarly, we found that clutch volume was correlated with predation risk and food availability, but not with vegetation cover. Regarding clutch size, the model with the lowest AIC score included predator species diversity, as well as maternal snout-vent length as explanatory factors. Similarly, the marginally better model for clutch volume included predation risk and maternal snout-vent length as independent variables.

Our analyses indicate that the main factor associated the evolution of different clutch sizes in island lizards is predator diversity. As the diversity of syntopic predators' decreases, so does clutch size, declining from an average of 4.12 eggs per clutch on Naxos Isl. to 1.47 eggs per clutch on predator-poor Glaronissi Isl. Similarly, predator species richness was the most important driver (albeit marginally) of clutch mass with the smallest clutch masses found on one off the most predator-poor islands (529 mm³, on Mikropsathoura). These results underscore the seminal importance that predation pressure plays for Lacertid lizards and dovetail with recent investigations that highlight the importance of predation as a generaldriving force for the evolution of island lizards. For example, intraspecific comparisons across numerous island populations have demonstrated that wall lizards on reduced-predation islands have both slower sprint speeds and tend to have relatively shorter legs (Semegen, 2018; Vervust et al., 2017). Furthermore, lizards on low-predation islands have downregulated tail autotomy (Savvides et al., 2017), stray further away from refugia (Cooper et al., 2000; Li et al., 2014) and are more apt to let potential predators approach before initiating escape behaviors (Li et al, 2014; Brock et al., 2015) relative to lizards on predator-rich mainland regions.

Previous studies have argued that food availability is a critical driver of reproductive output in vertebrates (Ballinger, 1977; Du, 2006; Mugabo et al., 2017). While in temperate lizards and in species living in strongly seasonal environments, increased food availability generally translates into larger clutches (Ballinger, 1977), in more tropical or aseasonal

environments it may instead result in more frequent clutches of the same size (Du, 2006). Although the Aegean Sea region is a strongly seasonal environment and *Podarcis* lizards tend to produce only one, or at most, two clutches annually, none of the models including food availability emerged as being best in explaining the observed clutch size variation. While there were significant correlations between food availability and both clutch size and clutch volume, the explanatory power was very small ($R_{adi}^2 = 0.024$ and 0.075 respectively). Part of this may be because the field methods did not allow us to adequately sample the preferred foods of the species. For example, P. erhardii appears to prefer to feed on larval Coleoptera and Orthoptera (Valakos, 1986), two groups of arthropods that are hard to sample either in pitfall or in sticky traps. Alternatively, it is known that clutch investment represents not just nutrients available during the reproductive season that are then shunted towards reproduction, but instead does also integrate stored lipids and is a reflection of the general longer-term nutritional status of an animal rather than recent nutritional income. Lastly, it is possible that the average clutch size and volume is a phylogenetically conservative trait that represents the long-term optimum for a population, rather than mapping tightly onto the local food availability in a given year. Either one of these may be the reason why we failed to detect a relationship between food availability and clutch size or volume.

Numerous earlier studies have shown that increased reproductive output, whether in mammals (McLean & Speakman, 2000), birds (Lee et al., 1996; Veasev et al., 2001) or reptiles (Seigel et al., 1987) incurs multiple costs that can affect future survival. Such costs stem from impaired thermoregulation (Shine, 1980) but especially from reduced locomotor performance which in turn impairs escaping ability from predators (Magnhagen, 1991; Brown & Shine, 2004). While the proximate drivers of reduced running speed in gravid females are complex (Olsson et al., 2000), much of the costs center on the need to escape rapidly while carrying additional offspring mass along (Seigel et al., 1987; Shine, 1980). As a result, gravid females tend to stay close to hiding places, and presence of sufficient cover and refugia can help females escape predation and perhaps thermoregulate more efficiently (Vervust et al., 2017). While P. erhardii generally prefers open areas it requires the presence of hiding places such as sclerophyllous *phrygana* and magui vegetation. As a matter of fact, lizard population density is related to the extent of sclerophyllous bush cover on an island (Foufopoulos et al. in prep.). As a result, we predicted that presence of cover in the form of evergreen shrubby vegetation and measured as an island's NDVI would be positively related to a population's reproductive investment. Instead, we found that there was no strong relationship between clutch size and NDVI. We also found that there was some support for a model incorporating NDVI to explain clutch volume, although this model was not the best (Table 2.).

In summary, when it comes to reproductive investment, wall lizard populations found in the Aegean Sea region constitute a textbook example of local adaptation showing different degrees of expression of the island syndrome depending on the extend of insularity of a particular population. Number of eggs produced by a female is not just a function of maternal SVL but is also strongly shaped by the richness of the resident predator community: lizard populations living on islands with the fewest predators showed a >64% reduction in clutch size, as well as corresponding reductions in clutch volume. At the same time, food availability appears to be significant factor for both clutch size and clutch volume: lizards living on islands with higher food availability have more eggs and larger clutches. Presence of vegetation appears to be positively associated with clutch size (but not clutch volume), presumably because more plant cover allows for slower-moving gravid females to avoid being prayed on and perhaps because of

thermoregulatory benefits. Traditional life history theory posits that the observed reductions in clutch size stem from a trade-off between clutch size and average egg (and therefore offspring) size. While in predator-rich environments selective pressures favor the production of large number of small-bodied offspring, high intraspecific competition among young lizards in low-predation, but high lizard density environments, is expected to favor investment in large-bodied offspring. Instead we find that low-predation populations produce *both* small clutches and eggs with relatively unchanged size. There are at least two possible explanations for this pattern. First, it is possible that low-predation islands are also low-productivity environments that do not provide resources for the production of large clutch numbers and volumes. Alternatively, it is possible that selection in low predation islands favors the evolution of long-lived life histories that produce small annual reproductive investment across many years. It is notable that these evolutionary patterns appear to have evolved several times and relatively rapidly. Some of the strongest inter-population differences occur on neighboring islands (e.g. on Naxos and Glaronissi which are <5km away from each other and have been separated for <5,000years) indicating that such differences can evolve quite quickly.

BIBLIOGRAPHY

- Abell, A. J., Cole, B. J., Reyes, R., & Wiernasz, D. C. (1999). Sexual selection on body size and shape in the western harvester ant, *Pogonomyrmex occidentalis* Cresson. *Evolution*, 53(2), 535-545.
- Adamopoulou, C., Valakos, E. D., & Pafilis, P. (1999). Summer diet of *Podarcis milensis, P. gaigeae* and *P. erhardii* (Sauria: Lacertidae). *Bonner Zoologische Beiträge*, 48(3/4), 275-282.
- Adler, G. H., & Levins, R. (1994). The island syndrome in rodent populations. *The Quarterly Review of Biology*, *69*(4), 473-490.
- Andrews, R. M. (1976). Growth rate in island and mainland anoline lizards. Copeia, 477-482.
- Angilletta, Jr, M. J., Niewiarowski, P. H., Dunham, A. E., Leaché, A. D., & Porter, W. P. (2004). Bergmann's clines in ectotherms: illustrating a life-history perspective with sceloporine lizards. *The American Naturalist*, 164(6), E168-E183.
- Arnold, E. N. (1987). Resource partition among lacertid lizards in southern Europe. *Journal of Zoology*, *1*(4), 739-782.
- Ballinger, R. E. (1977). Reproductive strategies: food availability as a source of proximal variation in a lizard. *Ecology*, *58*(3), 628-635.
- Ballinger, R. E. (1979). Intraspecific variation in demography and life history of the lizard, *Sceloporus jarrovi*, along an altitudinal gradient in southeastern Arizona. *Ecology*, 60(5), 901-909.
- Bauwens, D., & Thoen, C. (1981). Escape tactics and vulnerability to predation associated with reproduction in the lizard *Lacerta vivipara*. *The Journal of Animal Ecology*, 733-743.
- Bellard, C., Cassey, P., & Blackburn, T. M. (2016). Alien species as a driver of recent extinctions. *Biology Letters*, 12(2), 20150623. http://doi.org/10.1098/rsbl.2015.0623
- Blondel, J. (2000). Evolution and ecology of birds on islands: trends and prospects. *Vie et Milieu*, *50*(4), 205-220.
- Braña, F. (1993). Shifts in body temperature and escape behaviour of female *Podarcis muralis* during pregnancy. *Oikos*, 216-222.
- Brock, K. M., Donihue, C. M., & Pafilis, P. (2014). New records of frugivory and ovophagy in *Podarcis* (Lacertidae) lizards from East Mediterranean Islands. *North-Western Journal of Zoology*, 10(1), 223–225.
- Brock, K. M., Bednekoff, P. A., Pafilis, P., & Foufopoulos, J. (2015). Evolution of antipredator behavior in an island lizard species, *Podarcis erhardii* (Reptilia: Lacertidae): The sum of all

fears? Evolution, 69(1), 216-231. http://doi.org/10.1111/evo.12555

- Brown, G. P., & Shine, R. (2004). Effects of reproduction on the antipredator tactics of snakes (*Tropidonophis mairii*, Colubridae). *Behavioral Ecology and Sociobiology*, 56(3), 257-262.
- Case, T. J. (1975). Species numbers, density compensation, and colonizing ability of lizards on islands in the Gulf of California. *Ecology*, *56*(1), 3-18.
- Case, T.J., (1978). A general explanation for insular body size trends in terrestrial vertebrates. *Ecology*, 59(1), 1-18.
- Case, T. J., Bolger, D. T., & Richman, A. D. (1992). Reptilian Extinctions: The Last Ten Thousand Years. In: *Conservation Biology* (pp. 91–125). Boston, MA: Springer US. http://doi.org/10.1007/978-1-4684-6426-9_5
- Clegg, S. M., & Owens, P. F. (2002). The 'island rule'in birds: medium body size and its ecological explanation. *Proceedings of the Royal Society of London B: Biological Sciences*, *269*(1498), 1359-1365.
- Cooper, W. E., Vitt, L. J., Hedges, R., & Huey, R. B. (1990). Locomotor impairment and defense in gravid lizards (*Eumeces laticeps*): behavioral shift in activity may offset costs of reproduction in an active forager. *Behavioral Ecology and Sociobiology*, 27(3), 153-157.
- Cooper, W. E. (2000). Tradeoffs between predation risk and feeding in a lizard, the broadheaded skink (*Eumeces laticeps*). *Behaviour*, 137(9), 1175-1189.
- Cruz-Elizalde, R., & Ramírez-Bautista, A. (2016). Reproductive cycles and reproductive strategies among populations of the Rose-bellied Lizard *Sceloporus variabilis* (Squamata: Phrynosomatidae) from central Mexico. *Ecology and Evolution*, *6*(6), 1753-1768.
- Cruz-Elizalde, R., Ramírez-Bautista, A., Stephenson, B. P., Luja, V. H., & Hernández-Salinas, U. (2017). Variation in female reproduction between populations of the arboreal lizard *Urosaurus bicarinatus* (Squamata: Phrynosomatidae) from two different environments in Mexico. *Salamandra*, 53(3), 359-367.
- Díaz, J., Pérez-Tris, J., Tellería, J., Carbonell, R., & Santos, T. (2005). Reproductive investment of a Lacertid lizard in fragmented habitat. *Conservation Biology*, *19*(5), 1578-1585.
- Díaz, J. A., Iraeta, P., Verdú-Ricoy, J., Siliceo, I., & Salvador, A. (2012). Intraspecific variation of reproductive traits in a Mediterranean lizard: clutch, population, and lineage effects. *Evolutionary Biology*, *39*(1), 106-115.
- Du, W. G., Ji, X., Zhang, Y. P., Xu, X. F., & Shine, R. (2005). Identifying sources of variation in reproductive and life-history traits among five populations of a Chinese lizard (*Takydromus septentrionalis*, Lacertidae). *Biological Journal of the Linnean* Society, 85(4), 443-453.

- Du, W. G. (2006). Phenotypic plasticity in reproductive traits induced by food availability in a lacertid lizard, *Takydromus septentrionalis*. *Oikos*, *112*(2), 363-369.
- Dunham, A. E., & Miles, D. B. (1985). Patterns of covariation in life history traits of squamate reptiles: the effects of size and phylogeny reconsidered. *The American Naturalist*, 126(2), 231-257.
- Fielding, J., and N. Turland. (2008). *Flowers of Crete*. 2nd ed. Royal Botanic Gardens, Kew, London, Surrey, U.K.
- Flesness, N. R. (1989). Mammalian extinction rates: background to the black-footed ferret drama. pp. 3-9, in: Seal, U.S., Thorne, E.T., Bogan, M.A., and Anderson S.H. (eds.) *Conservation Biology of the Black-Footed Ferret*, Yale University Press, New Haven, CT, USA.
- Foufopoulos, J. (1999). Host-parasite interactions in the Mountain Spiny Lizard *Sceloporus jarrovi*. Ph.D. Thesis, University of Wisconsin 210pp.
- Foufopoulos, J., Kilpatrick, A. M., & Ives, A. R. (2011). Climate change and elevated extinction rates of reptiles from Mediterranean islands. *The American Naturalist*, *177*(1), 119–129. http://doi.org/10.1086/657624
- Galán, P. (2003). Reproductive characteristics of an insular population of the lizard *Podarcis hispanica* from Northwest Spain (Cies Islands, Galicia). *Copeia*, 2003(3), 657-665.
- Giorgi, F., & Lionello, P. (2008). Climate change projections for the Mediterranean region. *Global and Planetary Change*, 63(2), 90-104.
- Goltsman, M., Kruchenkova, E. P., Sergeev, S., Volodin, I., & Macdonald, D. W. (2005). 'Island syndrome'in a population of Arctic foxes (*Alopex lagopus*) from Mednyi Island. *Journal* of Zoology, 267(4), 405-418.
- Goodman, B. A., Hudson, S. C., Isaac, J. L., & Schwarzkopf, L. (2009). The evolution of body shape in response to habitat: is reproductive output reduced in flat lizards? *Evolution*, *63*(5), 1279-1291.
- Grove, A. T., & Rackham, O. (2003). *The Nature of Mediterranean Europe: An Ecological History*. Yale University Press.
- Haenel, G. (2011). Effects of habitat on clutch size of ornate tree lizards, Urosaurus ornatus. Western North American Naturalist, 247-256.
- Herrel, A., Huyghe, K., Vanhooydonck, B., Backeljau, T., Breugelmans, K., Grbac, I., ... Irschick, D. J. (2008). Rapid large-scale evolutionary divergence in morphology and performance associated with exploitation of a different dietary resource. *Proceedings of the*

National Academy of Sciences, *105*(12), 4792–4795. http://doi.org/10.1073/pnas.0711998105

- Hoy, S. R., Millon, A., Petty, S. J., Whitfield, D. P., & Lambin, X. (2016). Food availability and predation risk, rather than intrinsic attributes, are the main factors shaping the reproductive decisions of a long-lived predator. *Journal of Animal Ecology*, 85(4), 892-902.
- Huang, W. S. (2007). Ecology and reproductive patterns of the agamid lizard *Japalura swinhonis* on an east Asian island, with comments on the small clutch sizes of island lizards. *Zoological Science*, *24*(2), 181-188.
- Iverson, J. B., Higgins, H., Sirulnik, A., & Griffiths, C. (1997). Local and geographic variation in the reproductive biology of the snapping turtle (*Chelydra serpentina*). *Herpetologica*, 96-117.
- Jordan, M. A., & Snell, H. L. (2002). Life history trade-offs and phenotypic plasticity in the reproduction of Galapagos lava lizards (*Microlophus delanonis*). *Oecologia*, 130(1), 44-52.
- Kohn, D. D., & Walsh, D. M. (1994). Plant species richness--the effect of island size and habitat diversity. *Journal of Ecology*, 367-377.
- Lee, S. J., Witter, M. S., Cuthill, I. C., & Goldsmith, A. R. (1996). Reduction in escape performance as a cost of reproduction in gravid starlings, *Sturnus vulgaris*. *Proceedings of the Royal Society of London, B*, 263(1370), 619-623.
- Li, Y., Xu, F., Guo, Z., Liu, X., Jin, C., Wang, Y., & Wang, S. (2011). Reduced predator species richness drives the body gigantism of a frog species on the Zhoushan Archipelago in China. *Journal of Animal Ecology*, 80(1), 171–182. http://doi.org/10.1111/j.1365-2656.2010.01746.x
- Li, B., Belasen, A., Pafilis, P., Bednekoff, P., & Foufopoulos, J. (2014). Effects of feral cats on the evolution of anti-predator behaviours in island reptiles: insights from an ancient introduction. *Proceedings of the Royal Society of London, B*, 281(1788), 20140339.
- Lima, S. L., & Dill, L. M. (1990). Behavioral decisions made under the risk of predation: a review and prospectus. *Canadian Journal of Zoology*, 68(4), 619-640.
- Magnhagen, C. (1991). Predation risk as a cost of reproduction. *Trends in Ecology & Evolution*, 6(6), 183-186.
- Mayhew, W. W. (1963). Reproduction in the granite spiny lizard, *Sceloporus orcutti. Copeia*, 144-152.

- McLean, J. A., & Speakman, J. R. (2000). Morphological changes during postnatal growth and reproduction in the brown long-eared bat *Plecotus auritus*: implications for wing loading and predicted flight performance. *Journal of Natural History*, *34*(5), 773-791.
- Meiri, S. (2007). Size evolution in island lizards. *Global Ecology and Biogeography*, *16*(6), 702-708.
- Mesquita, D. O., & Colli, G. R. (2010). Life history patterns in South American tropical lizards. *Reproducción en reptiles: morfología, ecología y evolución*, 45-71.
- Mesquita, D. O., Costa, G. C., Colli, G. R., Costa, T. B., Shepard, D. B., Vitt, L. J., & Pianka, E. R. (2016). Life-history patterns of lizards of the world. *The American Naturalist*, 187(6), 689-705.
- Miles, D. B., Sinervo, B., & Frankino, W. A. (2000). Reproductive burden, locomotor performance, and the cost of reproduction in free ranging lizards. *Evolution*, 54(4), 1386-1395.
- Mugabo, M., Marquis, O., Perret, S., & Le Galliard, J. F. (2011). Direct and socially-mediated effects of food availability late in life on life-history variation in a short-lived lizard. *Oecologia*, *166*(4), 949-960.
- Niewiarowski, P. H., Angilletta, M. J., & Leaché, A. D. (2004). Phylogenetic comparative analysis of life-history variation among populations of the lizard *Sceloporus undulatus*: an example and prognosis. *Evolution*, *58*(3), 619-633.
- Novosolov, M., & Meiri, S. (2013). The effect of island type on lizard reproductive traits. *Journal of Biogeography*, 40(12), 2385-2395.
- Novosolov, M., Rodda, G. H., Feldman, A., Kadison, A. E., Dor, R., & Meiri, S. (2016). Power in numbers. Drivers of high population density in insular lizards. *Global Ecology and Biogeography*, 25(1), 87-95.
- Olsson, M. M. (2000). Locomotor impairment of gravid lizards: is the burden physical or physiological?. *Journal of Evolutionary Biology*, *13*, 263-268.
- Olsson, M., Shine, R., & Wapstra, E. (2001). Costs of reproduction in a lizard species: a comparison of observational and experimental data. *Oikos*, *93*(1), 121-125.
- Pafilis, P., Meiri, S., Foufopoulos, J., & Valakos, E. (2009). Intraspecific competition and high food availability are associated with insular gigantism in a lizard. *Naturwissenschaften*, 96(9), 1107-1113.
- Pafilis, P., Foufopoulos, J., Sagonas, K., Runemark, A., Svensson, E., & Valakos, E. D. (2011). Reproductive biology of insular reptiles: marine subsidies modulate expression of the "island syndrome". *Copeia*, 2011(4), 545-552.

- Polis, G. A., & Hurd, S. D. (1995). Extraordinarily high spider densities on islands: flow of energy from the marine to terrestrial food webs and the absence of predation. *Proceedings of the National Academy of Sciences*, 92(10), 4382-4386.
- Qualls, C. P., & Shine, R. (1995). Maternal body-volume as a constraint on reproductive output in lizards: evidence from the evolution of viviparity. *Oecologia*, *103*(1), 73-78.
- Reznick, D. (1985). Costs of reproduction: an evaluation of the empirical evidence. *Oikos*, 257-267.
- Reznick, D. (1992). Measuring the costs of reproduction. *Trends in Ecology and Evolution* 7:42-45.
- Rogers, L. E., Hinds, W. T., & Buschbom, R. L. (1976). A General Weight vs. Length Relationship for Insects. *Annals of the Entomological Society of America*, 69(2), 387–389. http://doi.org/10.1093/aesa/69.2.387
- Roitberg, E. S., Kuranova, V. N., Bulakhova, N. A., Orlova, V. F., Eplanova, G. V., Zinenko, O. I., ... & Yakovlev, V. A. (2013). Variation of reproductive traits and female body size in the most widely-ranging terrestrial reptile: testing the effects of reproductive mode, lineage, and climate. *Evolutionary Biology*, 40(3), 420-438.
- Runemark, A., Hansson, B., Pafilis, P., Valakos, E. D., & Svensson, E. I. (2010). Island biology and morphological divergence of the Skyros wall lizard *Podarcis gaigeae*: a combined role for local selection and genetic drift on color morph frequency divergence?. *BMC Evolutionary Biology*, 10(1), 269.
- Savvides, P., Stavrou, M., Pafilis, P., & Sfenthourakis, S. (2017). Tail autotomy affects bipedalism but not sprint performance in a cursorial Mediterranean lizard. *The Science of Nature*, 104(1-2), 3.
- Schaffer, W. M. (1974). Optimal reproductive effort in fluctuating environments. *The American Naturalist*, *108*(964), 783-790.
- Schwarzkopf, L. (1994) Measuring trade-offs: a review of studies of costs of reproduction in lizards. *Lizard Ecology: Historical and Experimental Perspectives* (eds L.J. Vitt & E.R. Pianka), pp. 7-30. Princeton University Press, Princeton, NJ, USA.
- Seigel, R. A., Huggins, M. M., & Ford, N. B. (1987). Reduction in locomotor ability as a cost of reproduction in gravid snakes. *Oecologia*, 73(4), 481-485.
- Semegen, S.L. (2018). Predation pressure as a determinant of locomotor performance: Lizards run slower on islands without predators. Unpublished MS. Thesis, Univ. of Michigan.

Shine, R. (1980). "Costs" of reproduction in reptiles. *Oecologia*, 46(1), 92-100.

- Shine, R. (2003). Locomotor speeds of gravid lizards: placing 'costs of reproduction' within an ecological context. *Functional Ecology*, *17*(4), 526-533.
- Shine, R. (2005). Life-history evolution in reptiles. *Annual Review of Ecology and Systematics*, *36*, 23-46.
- Shine, R., & Schwarzkopf, L. (1992). The evolution of reproductive effort in lizards and snakes. *Evolution*, *46*(1), 62-75.
- Siliceo, I., & Díaz, J. A. (2010). A comparative study of clutch size, range size, and the conservation status of island vs. mainland lacertid lizards. *Biological Conservation*, *143*(11), 2601-2608.
- Sinervo, B. (1990). The evolution of maternal investment in lizards: an experimental and comparative analysis of egg size and its effects on offspring performance. *Evolution*, *44*(2), 279-294.
- Sinervo, B., & Licht, P. (1991). Proximate constraints on the evolution of egg size, number, and total clutch mass in lizards. *Science*, *252*(5010), 1300-1302.
- Smith, C. C., & Fretwell, S. D. (1974). The optimal balance between size and number of offspring. *The American Naturalist*, 108(962), 499-506.
- Spatz, D. R., Zilliacus, K. M., Holmes, N. D., Butchart, S. H. M., Genovesi, P., Ceballos, G., ... Croll, D. A. (2017). Globally threatened vertebrates on islands with invasive species. *Science Advances*, 3(10). http://doi.org/10.1126/sciadv.1603080
- Stamps, J. A., & Buechner, M. (1985). The territorial defense hypothesis and the ecology of insular vertebrates. *The Quarterly Review of Biology*, 60(2), 155-181.
- Stearns, S. C. (1977). The evolution of life history traits: a critique of the theory and a review of the data. *Annual Review of Ecology and Systematics*, 8(1), 145-171.
- Stearns, S. C. (1992). *The Evolution of Life Histories*. Oxford; New York: Oxford University Press.
- Shure, D. J., & Phillips, D. L. (1991). Patch size of forest openings and arthropod populations. *Oecologia*, *86*(3), 325-334.
- Tinkle, D. W. (1969). The concept of reproductive effort and its relation to the evolution of life histories of lizards. *The American Naturalist*, *103*(933), 501-516.
- Tinkle, D. W., Wilbur, H. M., & Tilley, S. G. (1970). Evolutionary strategies in lizard reproduction. *Evolution*, 24(1), 55-74.

- Turkheimer, E., Haley, A., Waldron, M., d'Onofrio, B., & Gottesman, I. I. (2003). Socioeconomic status modifies heritability of IQ in young children. *Psychological Science*, 14(6), 623-628.
- Valakos, E. (1986). The feeding ecology of *Podarcis erhardii* (Reptilia-Lacertidae) in a main insular ecosystem. *Herpetological Journal*, 1(3), 118-121.
- Valakos, E., Pafilis, P., Sotiropoulos, K., Lymberakis, P., Maragou, P., & Foufopoulos, J. (2008). *The Amphibians and Reptiles of Greece*. Frankfurt, Germany: Chimaira.
- Van Damme, R., Bauwens, D., & Verheyen, R. F. (1989). Effect of relative clutch mass on sprint speed in the lizard *Lacerta vivipara*. *Journal of Herpetology*, *23*(4), 459-461.
- Veasey, J. S., Houston, D. C., & Metcalfe, N. B. (2001). A hidden cost of reproduction: the trade-off between clutch size and escape take-off speed in female zebra finches. *Journal of Animal Ecology*, *70*(1), 20-24.
- Vervust, B., Grbac, I., Damme, R. Van, Vervust, B., Grbac, I., & Damme, R. Van. (2017). Differences in morphology, performance and behaviour between recently diverged populations of *Podarcis sicula* mirror differences in predation pressure. *Oikos*, 116(8), 1343–1352.
- Vitt, L. J. (1981). Lizard reproduction: habitat specificity and constraints on relative clutch mass. *The American Naturalist*, 117(4), 506-514.
- Vitt, L. J., & Congdon, J. D. (1978). Body shape, reproductive effort, and relative clutch mass in lizards: resolution of a paradox. *The American Naturalist*, *112*(985), 595-608.
- Warne, R. W., Gilman, C. A., Garcia, D. A., & Wolf, B. O. (2012). Capital breeding and allocation to life-history demands are highly plastic in lizards. *The American Naturalist*, 180(1), 130-141.
- Werneck, F. D. P., Giugliano, L. G., Collevatti, R. G., & Colli, G. R. (2009). Phylogeny, biogeography and evolution of clutch size in South American lizards of the genus *Kentropyx* (Squamata: Teiidae). *Molecular Ecology*, 18(2), 262-278.
- Wikelski, M., Foufopoulos, J., Vargas, H., & Snell, H. (2004). Galápagos birds and diseases: invasive pathogens as threats for island species. *Ecology and Society*, 9(1).

APPENDICES

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1. R outputs of linear regression between island area and independent variables
Call:
lm(formula = log(datasum$PredatorSpeciesNumber) ~ log(datasum$IslandArea))
Residuals:
     Min
                   Median
                                        Max
              10
                                30
-0.36920 -0.12622 0.06443 0.14174 0.32167
Coefficients:
                       Estimate Std. Error t value Pr(>|t|)
(Intercept)
                        1.38070
                                   0.06256 22.07 4.40e-11 ***
log(datasum$IslandArea) 0.19048
                                   0.01308 14.56 5.47e-09 ***
___
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.2134 on 12 degrees of freedom
Multiple R-squared: 0.9464, Adjusted R-squared: 0.942
F-statistic: 212 on 1 and 12 DF, p-value: 5.466e-09
Call:
lm(formula = log(datasum$FoodBiomass) ~ log(datasum$IslandArea))
Residuals:
     Min
               10
                    Median
                                  30
                                          Max
-0.83461 -0.46454 -0.03503 0.44819 1.11602
Coefficients:
                         Estimate Std. Error t value Pr(>|t|)
(Intercept)
                          4.38242
                                     0.18285 23.967 1.67e-11 ***
log(datasum$IslandArea) 0.15160
                                     0.03824 3.964 0.00188 **
___
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.6237 on 12 degrees of freedom
Multiple R-squared: 0.567,
                                Adjusted R-squared: 0.5309
F-statistic: 15.72 on 1 and 12 DF, p-value: 0.001879
```

Call: lm(formula = datasum\$NDVI ~ datasum\$IslandArea) Residuals: Min 10 Median Max 3Q -0.17240 -0.10198 -0.03417 0.08694 0.23513 Coefficients: Estimate Std. Error t value Pr(>|t|) 0.2803522 0.0435775 6.433 3.24e-05 *** (Intercept) datasum\$IslandArea 0.0002752 0.0001057 2.605 0.023 * ___ Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 0.1357 on 12 degrees of freedom Multiple R-squared: 0.3611, Adjusted R-squared: 0.3079 F-statistic: 6.784 on 1 and 12 DF, p-value: 0.02303

2. R outputs of linear regressions between clutch size and independent variables

Call: lm(formula = data\$ClutchSize ~ data\$PredatorSpeciesNumber) Residuals: Min 10 Median 30 Max -1.9741 -0.5894 0.0259 0.6317 3.1990 Coefficients: Estimate Std. Error t value Pr(>|t|) 0.14093 10.050 < 2e-16 *** 1.41627 (Intercept) data\$PredatorSpeciesNumber 0.08655 0.01198 7.223 6.45e-12 *** Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 1.013 on 244 degrees of freedom Multiple R-squared: 0.1762, Adjusted R-squared: 0.1728 F-statistic: 52.18 on 1 and 244 DF, p-value: 6.449e-12

Call: lm(formula = data\$ClutchSize ~ data\$FoodBiomass) Residuals: Min 10 Median 3Q Max -1.9134 -0.8464 -0.1133 0.8083 3.4405 Coefficients: Estimate Std. Error t value Pr(>|t|) 1.7756044 0.1378230 12.883 < 2e-16 *** (Intercept) data\$FoodBiomass 0.0034153 0.0007495 4.557 8.22e-06 *** ____ Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 1.071 on 244 degrees of freedom Multiple R-squared: 0.07842, Adjusted R-squared: 0.07464 F-statistic: 20.76 on 1 and 244 DF, p-value: 8.218e-06 Call: lm(formula = data\$ClutchSize ~ data\$NDVI) Residuals: Min 10 Median 30 Max -1.8637 -0.9180 -0.1144 0.8392 3.4897 Coefficients: Estimate Std. Error t value Pr(>|t|) (Intercept) 0.1707 9.624 < 2e-16 *** 1.6429 0.4126 4.337 2.11e-05 *** data\$NDVI 1.7898 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 1.075 on 244 degrees of freedom Multiple R-squared: 0.07158, Adjusted R-squared: 0.06778 F-statistic: 18.81 on 1 and 244 DF, p-value: 2.111e-05

3. R outputs of linear regressions between clutch volume and independent variables

Call: lm(formula = data\$ClutchVolume ~ data\$PredatorSpeciesNumber) Residuals: 1Q Median Min 3Q Max -1117.4 -648.8 -159.5 431.8 3974.1 Coefficients: Estimate Std. Error t value Pr(>|t|) (Intercept) 400.42 121.17 3.305 0.00109 ** 10.30 4.322 2.26e-05 *** data\$PredatorSpeciesNumber 44.52 ___ Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 870.5 on 244 degrees of freedom Multiple R-squared: 0.0711, Adjusted R-squared: 0.0673 F-statistic: 18.68 on 1 and 244 DF, p-value: 2.255e-05 Call: lm(formula = data\$ClutchVolume ~ data\$FoodBiomass) Residuals: Min 10 Median Max 30 -1129.4 -651.7 -305.3 481.8 4179.7 Coefficients: Estimate Std. Error t value Pr(>|t|) (Intercept) 600.8882 114.5931 5.244 3.41e-07 *** data\$FoodBiomass 1.6590 0.6232 2.662 0.00828 ** Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 890.4 on 244 degrees of freedom Multiple R-squared: 0.02823, Adjusted R-squared: 0.02424 F-statistic: 7.087 on 1 and 244 DF, p-value: 0.008279

<u>4. R outputs of linear regressions between clutch size or clutch volume and maternal snout-vent length</u>

Call: lm(formula = data\$ClutchSize ~ data\$SVL) Residuals: Min 10 Median 30 Max -2.25142 -0.88268 -0.08376 0.73187 3.11732 Coefficients: Estimate Std. Error t value Pr(>|t|) (Intercept) -1.48067 0.79135 -1.871 0.0625 . data\$SVL 0.01274 4.822 2.5e-06 *** 0.06146 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 1.066 on 244 degrees of freedom Multiple R-squared: 0.087, Adjusted R-squared: 0.08326 F-statistic: 23.25 on 1 and 244 DF, p-value: 2.502e-06 Call: lm(formula = data\$ClutchVolume ~ data\$SVL) Residuals: Min 10 Median 30 Max -1202.4 -659.4 -347.7 577.5 3798.2 Coefficients: Estimate Std. Error t value Pr(>|t|) 659.85 -1.507 0.13313 (Intercept) -994.31 data\$SVL 10.63 2.830 0.00505 ** 30.07 ___ Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 888.8 on 244 degrees of freedom Multiple R-squared: 0.03177, Adjusted R-squared: 0.0278 F-statistic: 8.007 on 1 and 244 DF, p-value: 0.005048

5. R outputs of selected models to explain clutch size and clutch volume

```
Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
Formula: data$ClutchSize ~ data$PredatorSpeciesNumber + data$SVL + (1 |
    data$Location)
REML criterion at convergence: 704.7
Scaled residuals:
    Min
             1Q Median
                             3Q
                                   Max
-2.3326 -0.7350 0.1262 0.6095 2.7431
Random effects:
 Groups
                           Variance Std.Dev.
               Name
 data$Location (Intercept) 0.02392 0.1546
                           0.95159 0.9755
 Residual
Number of obs: 246, groups: data$Location, 14
Fixed effects:
                            Estimate Std. Error
                                                      df t value Pr(>|t|)
(Intercept)
                            -1.48402
                                       0.75195 168.12000 -1.974 0.050070 .
data$PredatorSpeciesNumber
                            0.07505
                                       0.01429 9.73000 5.253 0.000407 ***
                                                          3.966 0.000104 ***
data$SVL
                             0.04875
                                       0.01229 186.99000
____
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Correlation of Fixed Effects:
            (Intr) dt$PSN
dt$PrdtrSpN -0.009
data$SVL
            -0.978 -0.173
Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
Formula: data$ClutchVolume ~ data$PredatorSpeciesNumber + data$SVL + (1 |
    data$Location)
REML criterion at convergence: 3998.4
Scaled residuals:
    Min
             1Q Median
                             30
                                    Max
-1.4845 -0.7438 -0.2338 0.5773 4.3531
Random effects:
Groups
                           Variance Std.Dev.
               Name
 data$Location (Intercept) 14009 118.4
 Residual
                           735291
                                    857.5
Number of obs: 246, groups: data$Location, 14
Fixed effects:
                           Estimate Std. Error
                                                    df t value Pr(>ItI)
(Intercept)
                            -970.72
                                        656.43 183.32 -1.479 0.14091
data$PredatorSpeciesNumber
                              40.77
                                         12.08
                                                14.43
                                                         3.376 0.00436 **
data$SVL
                              22.68
                                         10.74 197.54
                                                         2.111 0.03600 *
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Correlation of Fixed Effects:
            (Intr) dt$PSN
dt$PrdtrSpN -0.001
data$SVL
            -0.979 -0.177
```

0. Arc tubles of thear mixed effect models of clutch size and thear models of clutch volume

Model	AICc	Δ AICc	Akaike Weight
CS ~ P + SVL + (1 Location)	714.703	0.000	0.687
$CS \sim NDVI + SVL + (1 Location)$	716.997	2.293	0.218
$CS \sim SVL + (1 Location)$	719.623	4.920	5.871x10 ⁻²
$CS \sim P + (1 Location)$	720.750	6.047	3.342x10 ⁻²
$CS \sim NDVI + (1 Location)$	727.220	12.517	1.315x10 ⁻³
$CS \sim B + SVL + (1 Location)$	727.674	12.971	1.049x10 ⁻³
$CS \sim B + (1 Location)$	739.570	24.867	2.737x10 ⁻⁶

Table 4. AIC for seven linear mixed effect models constructed to explain clutch size

CS = Clutch Size, P = Predator Richness, B = Biomass of Arthropods, SVL = maternal snoutvent length. Models were compared with their AICc values and their associated Akaike weightwhich is calculated by equation:

 $W_i(AIC) = exp(-0.5\Delta_i(AIC)) / \sum_{k=1}^k exp(-0.5\Delta_k(AIC)) \text{ (Turkheimer et al. 2003).}$

Model	AIC	Δ AIC	Akaike Weight
$CV \sim P + SVL$	4030.073	0	0.753
$CV \sim P$	4032.505	2.432	0.223
$CV \sim B + SVL$	4037.205	7.132	2.128x10 ⁻²
$CV \sim SVL$	4042.707	12.634	1.359x10 ⁻²
$CV \sim B$	4043.606	13.533	8.668x-10 ⁻⁴
$CV \sim NDVI + SVL$	4043.926	13.853	7.386x10- ⁴
$CV \sim NDVI$	4049.325	19.252	4.967x10 ⁻⁵

Table 5. AIC for seven linear models constructed to explain clutch volume

 $\overline{\text{CV} = \text{Clutch Volume}}$, $\overline{\text{P} = \text{Predator Richness}}$, $\overline{\text{B} = \text{Biomass of Arthropods}}$, $\overline{\text{SVL} = \text{maternal}}$ snout-vent length. Models were compared with their AICc values and their associated Akaike weight which is calculated by equation:

 $W_i(AIC) = exp(-0.5\Delta_i(AIC)) / \sum_{k=1}^k exp(-0.5\Delta_k(AIC))$ (Turkheimer et al. 2003).