

Biogeography of *Lacerta (Zootoca) vivipara*: reproductive mode and enzyme phenotypes in Bulgaria

Claude-Pierre Guillaume, Benoît Heulin and Vladimir Beshkov

Guillaume, C-P, Heulin, B and Beshkov, V 1997 Biogeography of *Lacerta (Zootoca) vivipara* reproductive mode and enzyme phenotypes in Bulgaria – *Ecography* 20 240–246

Lacerta (Zootoca) vivipara, has allopatric oviparous and viviparous populations viviparity is observed from central France and the British Isles to Scandinavia and Russia, while oviparity is restricted to northern Spain and southwestern France, i.e. the extreme southwestern part of the range. Recent observations in the Rila, Balkan, Vitocha, Pirin and Rhodopes mountains indicate that Bulgarian populations of *Lacerta (Zootoca) vivipara* are indeed viviparous. The electrophoretic study of allozymes and the estimation of genetic distances indicate that viviparous lizards from northwest and central France are more closely related to those of Bulgaria, than to the oviparous lizards of southwest France and northwest Spain. Variations in reproductive mode and allozymes are not directly related to geographic distances between populations, nor to their latitude. Populations located at the southwest limit of distribution are oviparous and exhibit alleles ATA⁻¹⁵⁰ or ATA⁻²⁰⁰, whereas, at a comparable latitude, the Bulgarian populations are viviparous and exhibit allele ATA⁻¹⁰⁰ characteristic of other distant viviparous populations. These findings underline the originality of the oviparous southwestern populations. They do not contradict our previous biogeographic scenario.

C-P Guillaume (ephebev@cru.univ-montp2.fr), E.P.H.E., Lab de Biogéographie et écologie, Univ de Montpellier II, Case courrier 094, F-34095 Montpellier Cedex 5, France – B Heulin, Station biologique de Pauvront, F-35380 Plélan-le-Grand, France – V Beshkov, Inst of Zoology, Bulgarian Academy of Sciences, 1 Tzar Osvoboditel, BU Sofia, Bulgaria

Oviparity is the ancestral reproductive mode of vertebrates (Angelini and Ghiara 1984, Yaron 1985, Wake 1989). Viviparity (live-bearing) has evolved from oviparity (egg-laying) independently in fish, amphibians, reptiles and mammals (Shine 1989). Blackburn (1982), in particular, has demonstrated that viviparity evolved on a minimum of 45 separate occasions in *Sauria* (lizards). Among current theories on the evolution of viviparity in reptiles the most widely accepted hypotheses suggest that viviparity may be an adaptation to the cold climatic conditions of high latitude and/or high altitude (see review in Fitch 1970, Packard et al 1977, Tinkle and Gibbons 1977, Shine and Bull 1979, Pilorge and Barbault 1981,

Angelini and Ghiara 1984, Shine 1985, Heulin et al 1991)

Because of the frequent apparition of viviparity in Squamata, one might expect to find key transitional species exhibiting both oviparous and viviparous reproductive modes. Such reproductive bimodality is a very rare phenomenon, reported reliably for only eleven species of lizards and snakes (ten in Shine 1985, more *Lacerta (Zootoca) vivipara*). Reproductive bimodality has been well documented in the old-world lizard *Lacerta (Zootoca) vivipara* which has allopatric oviparous (egg-laying + incubation in natura) and viviparous (gestation + parturition) populations (Lantz 1927, Braña 1986, Heulin 1988, 1990)

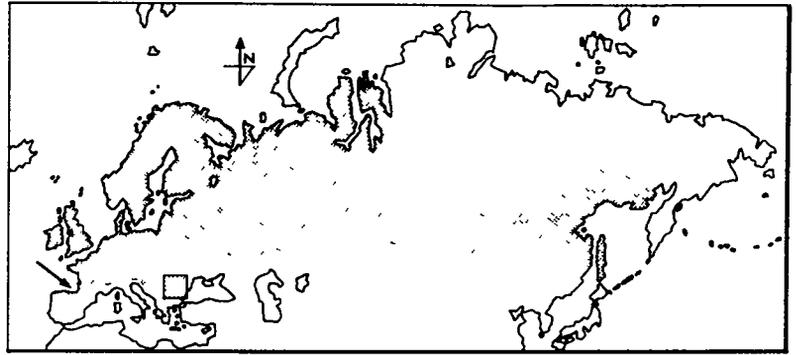
Accepted 7 October 1996

Copyright © ECOGRAPHY 1997

ISSN 0906-7590

Printed in Ireland – all rights reserved

Fig 1 Distribution area of *Lacerta (Zootoca) vivipara*. The arrow indicates the localisation of oviparous populations. The square indicates the localisation of Bulgarian populations.



Viviparous populations of *Lacerta (Zootoca) vivipara* are widely distributed from central France and the British Isles to Scandinavia and eastern Russia. Oviparous populations are isolated in the extreme southwestern part of the distribution area in the Pyrenees and Aquitaine in southwest France, and in northwest Spain (Figs 1 and 2) (Braña and Bea 1987, Heulin 1989, Heulin and Guillaume 1989).

Characteristics of viviparity (with persistence of non-functional oviparous structures), morphological and ecological resemblances, small genetic distances estimated from electrophoretic analyses, and successful experimental (laboratory) hybridisation all indicate that oviparous and viviparous strains are very closely related (Heulin et al 1989, 1992, 1993, Bea et al 1990, Arrayago et al 1996). This close relationship between the two strains also suggests that the emergence of viviparity is a recent event in this lizard species. Previously, we proposed the hypothesis stating that this recent emergence could have occurred during the cold climatic periods of the last quaternary Ice Age. In this case, populations (presumably still oviparous) would have retreated in two different directions, to the southwest and southeast. Viviparity could have evolved and been rapidly propagated in eastern populations while southwestern populations remained oviparous. After the glaciations, the newly evolved viviparous populations could have (re)colonised northwestern and northeastern countries, without any secondary contact with the residual oviparous populations isolated in the Pyreneo-Iberian refuge areas (Heulin et al 1993). The lack of secondary contact between oviparous and viviparous in France (Figs 1 and 2) could be the consequence of an ecological discontinuity. *L. (Zootoca) vivipara* prefers humid biotopes (wet heaths, peat bogs) and, because of climatic and edaphic conditions, such biotopes do not exist at present in the geographic area separating oviparous and viviparous populations in southern France (Chouard 1931, Heulin and Guillaume 1989, Heulin et al. 1993).

Our biogeographic hypothesis could well explain the recent evolution of viviparity and the existence of an isolated relictual form, i.e. the oviparous southwestern

populations. In Bulgaria, *L. (Zootoca) vivipara* is also considered a glacial relict; it is restricted to mountain areas (Western and Central Balkans, Vitocha, Rila, Western Rhodopes, Pirin) above 1200 m (Beshkov and Beron 1964, Beshkov 1984). A further interest of these isolated populations is that they are located on the southern limit of the species range, at the same latitude as Pyreneo-Iberic oviparous populations (Fig 1). Because of this geographic symmetry, the original oviparous strain could have retreated and remained unchanged in Bulgaria like in the Pyreneo-Iberian refuge. Alternatively, our asymmetric biogeographic scenario suggests that Bulgarian populations of *L. (Zootoca) vivipara* should be viviparous. So far, no information has been published on the reproductive mode of the species in Bulgaria. For these reasons we

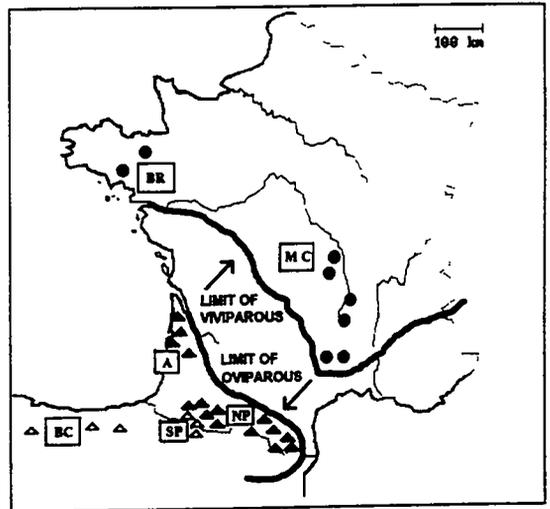


Fig 2 Origins of the oviparous and viviparous samples from Spain and France. Open triangles: oviparous from the Basque country and Cantabria in NW Spain (BC) and from the south slopes of the Pyrenees (SP). Black triangles: oviparous sample from the north slopes of the Pyrenees (NP) and from Aquitaine (A). Circles: viviparous sample from the Massif central mountains (MC) and Brittany (BR). Approximate northeast limit of oviparous and southwest limit of viviparous (after Heulin 1989).

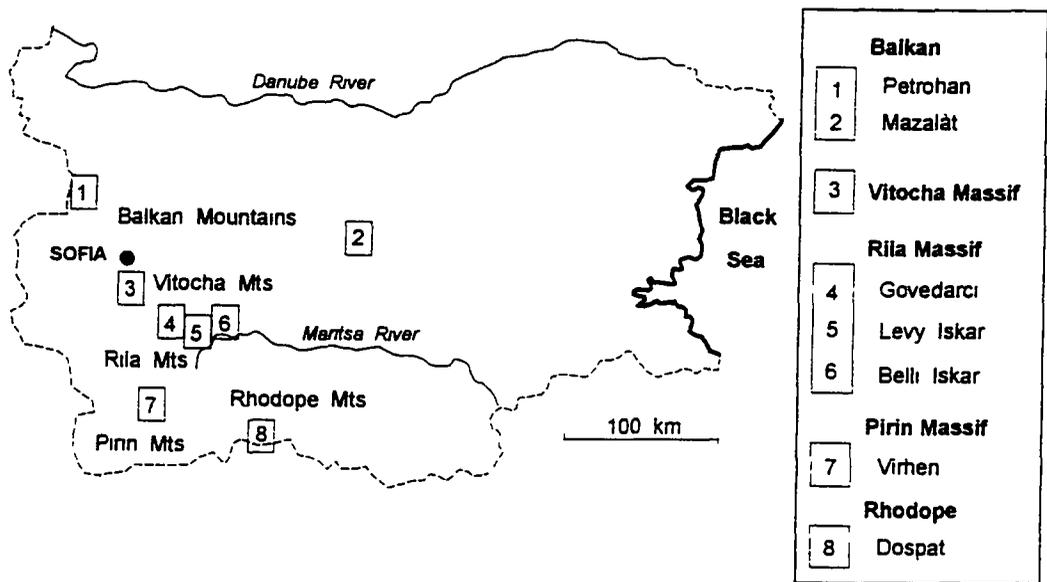


Fig 3 Localisation of the populations of *Lacerta (Zootoca) vivipara* studied in Bulgaria

set out to study the reproductive mode and the genetic (protein electrophoresis) characteristics of different populations of this species in Bulgaria (Fig 3)

Material and methods

Reproductive mode in Bulgarian populations of *L. vivipara*

We studied the reproductive mode of twenty-three gravid females captured in wild populations (stations 1, 3–7, Fig 3) and kept in laboratory terraria between 15 July and 15 August 1994: eight females from Petrohan (alt 1400–1700 m, lat 43°05' N, western Balkan), three females from the Vitocha mountain (alt 1850 m, lat 42°40' N), four females from Govedarci (alt 1300 m, lat 42°10' N, Rila mountains), two females from Levy-Iskar (alt. 1800 m, Rila), four females from Belli-Iskar (alt 1850 m, Rila), and two females from Vihren (alt 1950 m, lat 41°50' N, Pirin mountain)

Additional observations on the reproductive mode were available for a population located at Dospat (alt ca 1300 m, lat 41°40' N, station 2 in Fig. 3) in the western Rhodopes, and for another located at Mazalät (alt ca 1500 m, lat 42°50' N, station 8 in Fig 3) in the central Balkans (Beshkov unpubl and voucher specimens from the Bulgarian Museum of Natural History)

Electrophoretic comparisons between Bulgarian, French and Spanish lizards

All the animals were captured, transported and put to death with the authorisations of the competent Bulgarian,

Spanish and French administrations. For the genetic study of Bulgarian lizards, we used nine males and ten females: one male and one female from Vitocha, two males and one female from Belli-Iskar, one female from Levy-Iskar, two males and one female from Govedarci, three males and four females from Petrohan, one male and two females from Pirin (Fig 3)

The allozyme characteristics of this Bulgarian sample were compared to those of a viviparous sample from central and northwestern France (20 males, 16 females), and to those of an oviparous sample from southwest France and northwest Spain (36 males, 53 females). The majority of the lizards (77 out of 125) from the two latter samples had been used in a previous electrophoretic study (Bea et al 1990). The oviparous sample consisted of 1) 14 males and 11 females from three localities in the Basque country and Cantabria in NW Spain, (BC populations, lat 42°50' N, alt 600–1500 m), 2) seven males and 20 females from three localities on the south slopes of the Pyrenees (SP populations, lat 42°50' N, alt 1100–1900 m), 3) ten males and 19 females from 11 localities on the north slopes of the Pyrenees (NP populations, lat 42°30'–43°5' N, alt 380–2020 m), 4) four males and three females from four localities in Aquitaine (A populations, lat. 44°–45° N, alt 40–100 m) (Fig. 2). The viviparous sample consisted of 1) 12 males and nine females from six localities in the Massif Central mountains (MC populations, lat 43°40'–46° N, alt 970–1500 m), 2) eight males and seven females from two localities in Brittany (BR populations, lat 48° N, alt 100–150 m) (Fig 2)

Liver and muscle tissues were excised and stored at –30°C awaiting preparation for enzyme extraction. Starch gel electrophoresis techniques (Pasteur et al.

Table 1 Protein trivial names and abbreviations, International code numbers of the Enzyme Commission, number of loci, and electrophoretic systems used a Tris-borate-citrate pH 8.7 ("Poulik"), b Tris-citrate pH 8.0, c Tris-citrate pH 6.7 with NaDP

Protein (code)	E C No	N loci	Electrophoretic system
Lactate dehydrogenase (LDH)	1 1 1 27	2	a
Malate dehydrogenase (MDH)	1 1 1 37	1	b
Malic enzyme (ME)	1 1 1 40	1	c
Phosphogluconate dehydrogenase (PGD)	1 1 1 43	1	b
Nucleoside phosphorylase (NP)	2 4 2 1	1	a
Aspartate transaminase (ATA)	2 6 1 1	2	b
Adenylate kinase (AK)	2 7 4 3	1	c
Phosphoglycerate mutase (PGM)	2 7 5 1	1	a
Mannose-phosphate isomerase (MPI)	5 3 1 8	1	c
Glucose-phosphate isomerase (GPI)	5 3 1 9	1	a

1988) were applied to 12 presumed loci (LDH-1, LDH-2, MDH, ME, PGD, NP, ATA⁻, ATA⁺, AK, PGM, MPI, GPI) (see Bea et al 1990) (Table 1)

Factorial analyses of correspondences (FAC) and calculation of genetic distance index (Nei 1972, 1978) were used to compare populations from Bulgaria with the viviparous sample from France, and with the oviparous sample from southwest France and northwest Spain. Because of geographic variations in the allelic frequencies of MPI and NP loci (see Bea et al 1990), we divided the oviparous sample into two subsets (BC + SP = open triangles and NP + A = black triangles in Fig. 2)

FAC were computed in disjunctive allelic coding with "BIOMEKO" software (Lebreton et al 1990)

Results

Reproductive mode

All the Bulgarian lizards (from all localities) exhibited viviparous reproduction. Parturition occurred between 21 July and 8 August and the mean fecundity was six youngs per female ($n = 23$, $SD = 1.7$). Moreover, viviparous reproduction had previously been observed in a population located at Dospat (alt 1300 m, station 2 in Fig. 2) in the western Rhodopes (Beshkov unpubl.), and another was confirmed at Mazalat (station 8 in Fig. 2) from specimens from the Bulgarian Museum of Natural History

Protein electrophoresis

The relative frequency of alleles are presented in Table 2. From a qualitative point of view, allele ATA⁻¹⁰⁰ has previously been observed to be characteristic of viviparous populations from central and western France, whereas alleles ATA⁻¹⁵⁰ and ATA⁻²⁰⁰ are characteristic of oviparous populations from southwest France and northwest Spain (Bea et al 1990). The

present study demonstrates that allele ATA⁻¹⁰⁰ also is characteristic of the viviparous populations of Bulgaria (Table 2). This result confirms the presence, for the ATA⁻ locus, of diagnostic alleles separating oviparous from viviparous lizards in all the populations of *L. Zootoca vivipara* studied so far.

With regard to the MPI locus, some alleles have been shown to be sex-linked in both viviparous females (MPI¹¹⁰) (Goux and Pasteur 1986, Salvidio et al 1990) and oviparous females (MPI¹¹⁰ for the majority of "French" individuals and MPI⁹⁰ for the others and for all the "Spanish" ones) (Bea et al 1990). Our results indicate that the same phenomenon exists in viviparous Bulgarian populations.

The projection on the principal plane of the FAC emphasises the importance of loci ATA⁻ and NP for separating the animals into distinct subsets. Some alleles of loci ME, PGD, and GPI also contribute, to a lesser extent, to separating groups (Fig. 4). At the left of the first axis, we can see oviparous lizards from Spain and the south slopes of the Pyrenees (with ATA⁻¹⁵⁰ and ATA⁻²⁰⁰) while viviparous animals (monomorphic for the ATA⁻¹⁰⁰ allele) are at the right. At the left, at the top of the second axis, we find oviparous French and Spanish lizards with allele MPI⁹⁰ and a high frequency of allele NP¹⁰⁰, while oviparous French animals with allele MPI¹¹⁰ are at the bottom. At the right, viviparous French and Bulgarian lizards (with allele MPI¹¹⁰ or MPI¹²⁰, and higher frequency of allele NP¹⁵⁰) are mixed in the centre.

Finally it can be observed that viviparous lizards from Bulgaria are relatively close to viviparous French lizards and distant from other subsets. Furthermore we estimated Nei's genetic distances for all the animals in each subset (Bulgarian viviparous, French viviparous, NP + A oviparous, SP + BC oviparous). These distances (Table 3) also demonstrate that viviparous Bulgarian populations are more similar to viviparous French populations than they are to oviparous ones. The overall Nei's mean genetic distance (D_N) between viviparous (Bulgarian + French) and oviparous (French + Spanish) lizards is 0.102.

Table 2 Relative frequencies of observed allozymes in *Lacerta (Zootoca) vivipara* samples. Oviparous samples OV1 = from the Basque country and Cantabria (BC) and south slopes of the Pyrenees (SP), OV2 = from the north slopes of the Pyrenees (NP) and from Aquitaine (A). Viviparous samples VF = from France, VB = from Bulgaria.

Enzyme locus	Alleles	Oviparous samples				Viviparous samples	
		OV1	OV2	OT (OV1 + OV2)	VF	VB	VT (VF + VB)
ME	90	-	0.04	0.02	0.05	0.16	0.10
	100	0.26	0.22	0.24	0.43	0.50	0.46
	105	0.18	0.12	0.16	0.20	0.08	0.15
	110	0.20	0.27	0.23	0.07	0.18	0.12
	120	0.24	0.24	0.24	0.25	0.08	0.17
	130	0.12	0.11	0.11	-	-	-
PGD	80	0.06	0.03	0.05	0.17	0.03	0.10
	100	0.91	0.64	0.80	0.73	0.97	0.85
	120	0.03	0.33	0.15	0.10	-	0.05
NP	100	0.55	0.76	0.64	0.85	0.39	0.67
	150	0.45	0.24	0.36	0.15	0.61	0.33
ATA ⁺	100	1.00	0.97	0.99	1.00	0.84	0.94
	150	-	0.03	0.01	-	0.16	0.06
ATA ⁻	100	-	-	-	1.00	1.00	1.00
	150	0.42	0.53	0.47	-	-	-
	200	0.58	0.47	0.53	-	-	-
AK	100	0.95	0.84	0.90	0.83	1.00	0.90
	150	-	0.07	0.03	0.17	-	0.10
	200	0.05	0.09	0.07	-	-	-
PGM	60	0.02	-	0.01	-	-	-
	80	0.12	-	0.07	0.02	-	0.01
	100	0.80	0.92	0.85	0.96	0.82	0.91
	120	0.06	0.08	0.07	0.02	0.18	0.08
MPI	90	0.29	-	0.17	-	-	-
	100	0.71	0.70	0.71	0.74	0.71	0.73
	110	-	0.27	0.11	0.19	0.19	0.19
	120	-	0.03	0.01	0.07	0.10	0.08
GPI	60	0.03	-	0.02	0.03	-	0.02
	80	0.12	0.18	0.14	0.09	0.13	0.11
	100	0.85	0.79	0.83	0.88	0.87	0.87
	120	-	0.03	0.01	-	-	-

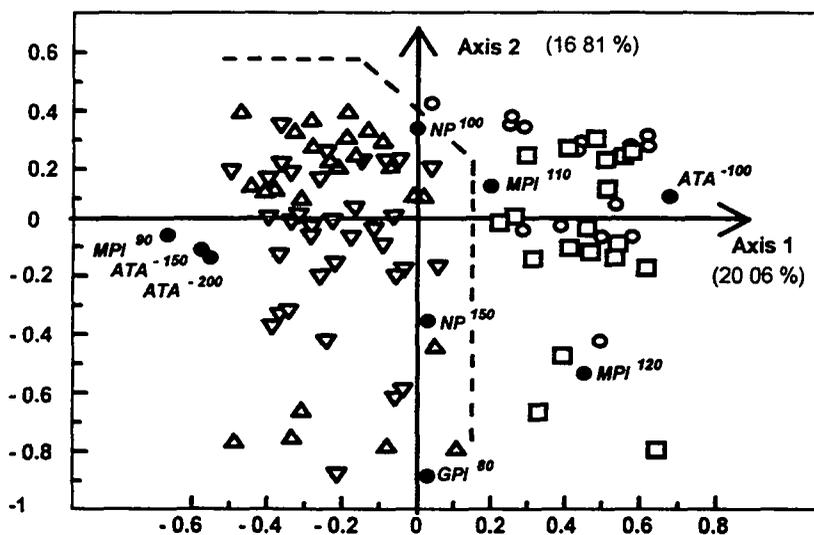


Fig 4 Projection of sampled lizards and of preponderant alleles on the principal plane (axes 1 and 2) of a FAC (factorial analysis of correspondences) (allelic coding) (More than one individual may be plotted in the same place). Squares = viviparous lizards from Bulgaria. Open circles = viviparous lizards from France. Upward triangles = oviparous lizards from north Pyrenean slope and Aquitaine. Downward triangles = oviparous lizards from south Pyrenean slope and from the Basque country and Cantabria. Filled circles = position of the preponderant alleles on the two first axes of the FAC. Dotted line = separation between oviparous and viviparous populations.

Table 3 Genetic distances (Ner's index, Nei 1978) between the various subsets of oviparous (O) and viviparous (V) populations of *Lacerta (Zootoca) vivipara*

	VB	OV1	OV2 (NP + A)	OT (OV1 + OV2)
VF (France BR + MC)	0 056	0 130	0 116	*
VB (Bulgaria St [1] + [3 - > 7])	*	0 121	0 150	*
OV1 (BC + SP)	*	*	0 039	*
VT (VF + VB)	*	*	*	0 102

These small genetic distances are similar, or inferior to distances between subspecies in other Lacertid lizards (Gorman et al 1975, Guillaume and Lanza 1982)

Discussion

Our studies on the reproductive mode and on the allozyme characteristics indicate that lizards from northwest and central France are more closely related to those of Bulgaria, than they are to the oviparous lizards from southwest France and northwest Spain. On the one hand, the genetic distance between viviparous French and Bulgarian lizards (0.056) is ca half the genetic distance between viviparous and oviparous French lizards (0.102). This clearly contrasts with geographic distances ca 1700 km between French and Bulgarian viviparous populations, and only 100 km between French oviparous and viviparous populations (i.e. distances between South MC viviparous populations and NP oviparous populations in Fig. 2). Furthermore, our observations also clearly indicate that, despite their proximity to the southern limit of distribution, Bulgarian populations of *L. (Zootoca) vivipara* are really viviparous.

Finally it can be emphasised that the variations in reproductive mode and allozymes are not directly related to the geographic distances between populations, nor to their latitude. Our study clearly underlines an asymmetrical distribution pattern: populations located at the southwest limit of the range are oviparous and exhibit alleles ATA⁻¹⁵⁰ or ATA⁻²⁰⁰ whereas, at a comparable latitude, the Bulgarian populations are viviparous and exhibit allele ATA⁻¹⁰⁰ characteristic of other distant viviparous populations. This finding supports, or at least does not contradict, our biogeographic scenario (cf. Introduction) predicting that viviparity could have evolved (first appeared) and been propagated in eastern populations, while oviparity remained unchanged in an isolated southwestern refuge (Heulin et al 1993). Unfortunately, the location where viviparity first appeared remains unknown. The hypothesis that this evolution could have occurred somewhere between the southern Carpathes and the southern Ural is not excluded. It remains to be studied using genetic comparisons (protein electrophoresis, nuclear

and mitochondrial DNA) in a larger number of viviparous populations from the whole distribution area.

To conclude, it seems interesting to compare our results with those of Alcobendas et al (1996) concerning the Urodele *Salamandra salamandra*. This species also exhibits the same two reproductive modes. Viviparous animals are localised in the Pyrenees and Cantabrian mountains (oviparous in lizards), and "[] the process is complicated by a complex sequence of secondary contacts []" between oviparous and viviparous populations resulting in the observation of populations with mixed reproductive mode. Dorsal colour patterns and body size allow the differentiation of large blotched oviparous "subspecies" from small striped viviparous and "mixed" ones (although we are unable to demonstrate any diagnostic morphologic feature between oviparous and viviparous *L. (Zootoca) vivipara*). Alcobendas et al (1996), studying the geographic variability in allozymes (34 loci), did not find any diagnostic alleles (whereas ATA⁻ is a diagnostic locus between oviparous and viviparous lizards).

In spite of these differences, for both species 1) the mean genetic distances among populations sampled (\bar{D}_N , varying from 0.050 to 0.199 in *S. salamandra* and from 0.039 to 0.150 [Table 3] in *L. (Zootoca) vivipara*) fall within the intraspecific ranges reported for other comparable animals. Oviparous and viviparous populations are undoubtedly conspecific. 2) There is no direct relationship between genetic distances and geographic distances. Oviparous populations are closer to each other than to viviparous, and conversely. 3) Phylogenetic hypotheses are fairly comparable: viviparity having evolved once in the group and occurring as an intraspecific variation, followed by biogeographical and/or ecological isolation of one form, and radiation of the other, the evolution of the Pyrenean and Cantabrian populations being related to the history of glacial activity in the region.

What would occur in oviparous *L. (Zootoca) vivipara* if a secondary contact with the viviparous took place? *Salamandra salamandra* could be a predictive model as, at present, in Squamata "no single population of any species shows both oviparity and viviparity [.] When a species exhibits both modes of reproduction, the egg-layers and live-bearers inhabit different geographic areas" (Shine 1985).

Acknowledgements – Financial support was provided by the French National Centre of Scientific Research CNRS-UA 1853, by funds from the French Ministry of Environment (EGPN-Reseau Metapopulation), and by the French and Bulgarian program "observatoire de Moussala" (S P Carbonell) Studies of allozymes were performed in the laboratory of J Deunff (Univ of Rennes), and in the laboratory of Fr Bonhomme (Univ of Montpellier II) The authors thank P Arntzen for his comments and suggestions and A Cloarec and E Guillosson for corrections to the manuscript

References

- Alcobendas, M., Dopazo, H and Alberch, P 1996 Geographic variation in allozymes of populations of *Salamandra salamandra* (Amphibia Urodela) exhibiting distinct reproductive modes – *J Evol Biol* 9 83–102
- Angelini, F and Ghiara, G 1984 Reproductive modes and strategies in vertebrate evolution – *Boll Zool* 51 121–203
- Arrayago, M-J, Bea, A and Heulin, B 1996 Hybridization experiment between oviparous and viviparous strains of *Lacerta vivipara* a new insight into viviparity evolution in reptiles – *Herpetologica* 52 333–342
- Beshkov, V A 1984 The effect of the Balkan range on the distribution of the herpetofauna in Bulgaria – *Acta Zool Bulg* 25 9–15
- and Beron, P 1964 Catalogue et bibliographie des Amphibiens et Reptiles en Bulgarie – Ed Acad Sci Bulg, Inst Zool Mus Sofia
- Bea, A, Guillaume, C-P, Arrayago, M-J, Heulin, B and Pasteur, G 1990 Phenotypes enzymatiques de *Lacerta (Zootoca) vivipara* premieres donnees comparatives entre populations ovipares et vivipares de cette espece – *C R Acad Sci Paris* 310 (ser III) 237–243
- Blackburn, D 1982 Evolutionary origins of viviparity in the Reptilia I Sauria – *Amphibia-Reptilia* 3 185–205
- Braña, F 1986 Ciclo reproductor y oviparismo de *Lacerta vivipara* en la cordillera Cantabrica – *Rev Española Herpetol* 1 274–291
- and Bea, A 1987 Bimodalite de reproduction chez *Lacerta vivipara* – *Bull Soc Herp France* 44 1–5
- Chouard, P 1931 Documents cartographiques sur les tourbières actuelles et préhistoriques de France – *Comp Rend Congr Int Geogr, Armand Colin, Paris*, pp 1–27
- Fitch, H S 1970 Reproductive cycles in lizards and snakes – *Univ Kans Mus Nat Hist* 52 1–247
- Gorman, G C, Soulé, M, Yang, S Y and Nevo, E 1975 Evolutionary genetics of insular adriatic lizards – *Evolution* 29 52–71
- Goux, J M and Pasteur, G 1986 A sex-linked enzyme in a reptile – association with a recent centric fusion in the common lizard – *Genet Res Camb* 48 21–25
- Guillaume, C-P and Lanza, B 1982 Comparaisons électrophorétiques de quelques especes de Lacertidés méditerranéens, genera *Podarcis* et *Archaeolacerta* – *Amphibia-Reptilia* 4 361–375
- Heulin, B 1988 Données nouvelles sur les populations ovipares de *Lacerta vivipara* – *C.R Acad Sci Paris* 306 (ser III) 63–68
- 1989 *Lacerta vivipara* – In Castanet, J and Guyetant, R (eds), Atlas de répartition des amphibiens et reptiles de France Soc Herp de France, Paris, pp 132–133
- 1990 Etude comparative de la membrane coquilliere chez les souches ovipares et vivipares de *Lacerta vivipara* – *Can J Zool* 68 1015–1019
- and Guillaume, C-P 1989. Extension géographique des populations ovipares de *Lacerta vivipara* – *Rev Ecol, Terre Vie* 44 39–45
- Arrayago, M-J and Bea, A 1989 Experience d'hybridation entre les souches ovipares et vivipares de *Lacerta vivipara* – *C R Acad Sci Paris* 308 (ser III) 341–346
- Oseneck, K and Lebouvier, M 1991 Timing of embryonic development and birth dates in oviparous and viviparous strains of *Lacerta vivipara* testing the predictions of an evolutionary hypothesis – *Acta Oecol Oecol Gen* 12 517–528
- Arrayago, M-J, Bea, A and Braña, F 1992 Caractéristiques de la coquille des oeufs chez la souche hybride (ovipare × vivipare) du lézard *Lacerta vivipara* – *Can J Zool* 70 2242–2246
- Guillaume, C-P, Bea, A and Arrayago, M-J 1993 Interprétation biogéographique de la bimodalité de reproduction du lézard *Lacerta vivipara* un modèle pour l'étude de l'évolution de la viviparité – *Biogeographica* 69 3–14
- Lantz, L A 1927 Quelques observations nouvelles sur l'herpétologie des Pyrénées centrales – *Rev Hist Nat Appliq* 8 54–61
- Lebreton, J D, Roux, M, Banco, G and Bacou, A M 1990 BIOMEKO (Biometrie-Ecologie) software, ver 3.9 – C E F E-C N R S Montpellier, France
- Nei, M 1972 Genetic distance between populations – *Am Nat* 106 283–292
- 1978 Estimations of average heterozygosity and genetic distance from a small number of individuals – *Genetics* 89 583–590
- Packard, G C, Tracy, C R and Roth, J-J 1977 The physiological ecology of reptilian eggs and embryos, and the evolution of viviparity within the class reptilia – *Biol Rev* 52 71–105
- Pasteur, N, Pasteur, G, Bonhomme, Fr, Catalan, J and Britton-Davidian, J 1988 Practical isozyme genetics – Ellis Horwood/John Wiley and Sons
- Pilorge, T and Barbault, R 1981 La viviparité chez les lézards evolution et adaptation – *Acta Oecol Oecol Gen* 2 387–397
- Salvidio, S, Pasteur, G, Heulin, B, Böhme, W, Kupriyanova, L and Guillaume, C-P 1990 Natural selection and geographical variation in a known sex-linked gene of the common lizard in Europe Implications for chromosomal evolution – *Heredity* 64 131–138
- Shine, R 1985 The evolution of viviparity in reptiles an ecological analysis – In Gans, C and Billet, F (eds), *Biology of the Reptilia*, Vol 15 John Wiley and Sons, pp 605–694
- 1989 Ecological influences on the evolution of vertebrate viviparity – In Wake, D B and Roth, G (eds), *Integration and evolution in vertebrates* John Wiley and Sons, pp 263–278
- and Bull, J J 1979 The evolution of live-bearing in lizards and snakes – *Am Nat* 113 905–923
- Tinkle, D W and Gibbons, J W 1977 The distribution and evolution of viviparity in Reptiles – *Misc Publ Univ Michigan* 154 1–55
- Wake, M H 1989 Phylogenesis of direct development and viviparity in vertebrates – In Wake, D B and Roth, G (eds), *Integration and evolution in vertebrates* John Wiley and Sons, pp 235–250
- Yaron, Z 1985 Reptilian placentation and gestation structure, function, and endocrine control – In Gans, C and Billet, F (eds), *Biology of the Reptilia*, Vol 15 John Wiley and Sons, pp 528–603

This document is a scanned copy of a printed document. No warranty is given about the accuracy of the copy. Users should refer to the original published version of the material.